

Is maternal serum total hCG a marker of trisomy 21 in the first trimester of pregnancy?

Kevin Spencer^{1*}, Esther Berry², Jennifer A. Crossley², David A. Aitken² and Kypros H. Nicolaides³

¹Endocrine Unit, Clinical Biochemistry Department, Harold Wood Hospital, Gubbins Lane, Romford, Essex, RM3 0BE, U.K.

²Institute of Medical Genetics, Yorkhill, Glasgow G3 8SJ, U.K.

³Harris Birthright Research Centre for Fetal Medicine, Kings College Hospital, Denmark Hill, London, U.K.

In a study of 130 first trimester cases of trisomy 21 and 959 controls we have shown that the median MoM for alpha-fetoprotein (AFP) is lower (0.82) and that for total human chorionic gonadotrophin (hCG) is higher (1.31) than in the control group. For AFP 15.3% of cases were below the 5th centile and for total hCG 19.8% were above the 95th centile. The median shift observed for AFP and total hCG is poorer than that for pregnancy associated plasma protein-A (PAPP-A) or free β -hCG and together with maternal age, AFP and total hCG could only be expected to detect 40% of cases. In combination with PAPP-A, total hCG would identify 52% of cases, somewhat less than the 67% observed with free β -hCG and PAPP-A. However, we have demonstrated for total hCG a significant temporal change in median MoM with gestational age. Before 70 days the median MoM was less than 0.5, between 70 and 83 days this increased to 1.13, and between 84 and 97 days this increased to 1.52. This median shift has significant implications for interpreting previous studies and even more significant implications for detection rates. When population parameters specific to the gestational age in question are used, detection rates with total hCG and PAPP-A increase from 47% at 70–83 days to 60% at 84–97 days. This observation explains much of the confusion around total hCG in the first trimester and shows the importance of selecting analyte pairs and population parameters appropriate to the time in gestation when screening is performed. Copyright © 2000 John Wiley & Sons, Ltd.

INTRODUCTION

Screening for trisomy 21 in the second trimester by the measurement of maternal serum biochemical markers has become an accepted part of obstetric practice in many countries (Cuckle *et al.*, 1995; Palomaki *et al.*, 1997). In routine practice a combination of maternal age with serum alpha-fetoprotein (AFP) and either free β -human chorionic gonadotrophin (hCG), with or without unconjugated oestriol can identify approximately 65% of cases for a 5% false positive rate (Crossley *et al.*, 1994; Wald *et al.*, 1992; Goodburn *et al.*, 1994; Macri and Spencer, 1996; Spencer, 1999a). In the first trimester of pregnancy much interest is focused on the use of fetal nuchal translucency (NT) thickness in combination with appropriate biochemical markers. Of the biochemical markers investigated, AFP and total hCG appear to be weak markers (Aitken *et al.*, 1993) in the first trimester, while only free β -hCG (Spencer *et al.*, 1992; Spencer, 1997) and pregnancy associated plasma protein-A (PAPP-A) have been shown to be of any value (Brambati *et al.*, 1991; Spencer *et al.*, 1994; Wald *et al.*, 1996). Large-scale studies (Spencer *et al.*, 1999a) have shown that detection rates of 89–90% can be achieved at a 5% false positive rate when ultrasound, maternal serum PAPP-A and free β -hCG are combined together at

10–14 weeks of gestation. Furthermore, the use of rapid immunodiagnostic technology has allowed the delivery of such screening within a 1 h time frame, leading to the development of a one-stop clinic for assessment of risk of fetal abnormality (OSCAR) (Spencer, 1999b).

Despite a general consensus that total hCG is of little value in screening for trisomy 21 in the first trimester (Wald *et al.*, 1997), sporadic reports in small series continue to claim that total hCG is of value at this time (Forest *et al.*, 1995; Haddow *et al.*, 1998; Casals *et al.*, 1999). We have therefore sought to clarify this situation by analysing AFP and total hCG in a large series of trisomy 21 cases, comparing these markers with other first trimester marker data largely from our previous study (Spencer *et al.*, 1999a).

METHODS

The study population was derived from three groups of women. The first comprised women with singleton pregnancies who were referred to the Harris Birthright Centre for fetal karyotyping, because screening by a combination of maternal age and fetal NT at 10–14 weeks in their hospital identified these patients as being at high risk for trisomy 21 (Snijders *et al.*, 1998). The second group comprised self-referred women for assessment of risk. Blood samples were collected from women at the time of the scan and the serum was aliquoted and stored at -20°C prior to blinded retrospective analysis. Gestational age was determined

*Correspondence to: K. Spencer, Endocrine Unit, Department of Clinical Biochemistry, Harold Wood Hospital, Gubbins Lane, Romford, Essex, RM3 0BE, U.K.
E-mail: Kevin_Spencer@Compuserve.com

by measurement of fetal crown–rump length (CRL). Pregnancy outcome was ascertained in all women. The 90 trisomy 21 cases formed part of a previous study of the markers free β -hCG and PAPP-A (Spencer *et al.*, 1999a). To supplement these a third group of 40 samples from cases of trisomy 21 were obtained from the Glasgow centre as part of an ongoing study (Berry *et al.*, 1997), which included the Combined Ultrasound and Biochemistry study (CUBS) in Scotland. In this series, total hCG, AFP, PAPP-A and free β -hCG were all measured at the same time.

To establish suitable reference data a control group of samples spreading the gestational ranges 10–14 weeks were obtained from samples collected as part of routine first trimester screening in the OSCAR clinic (Spencer, 1999b) at Harold Wood in patients with a normal pregnancy outcome. All samples were analysed for PAPP-A and free β -hCG within 30 min of sample collection and samples were then subsequently frozen as aliquots at -20°C . Pregnancy outcome was ascertained in all women. To supplement these, a further 90 control samples in the gestational period 6–9 weeks were obtained from the Glasgow centre.

In total, serum samples from 130 pregnancies affected by trisomy 21 were available for biochemical analysis. A control group of 959 cases resulting in the birth of an unaffected baby were used to establish median and reference data. Table 1 shows the characteristics of the trisomy 21 population and also that for the control population.

Maternal serum AFP and total hCG were measured over a period of three days on the Kryptor analyser—a random access immunoassay analyser using time-resolved amplified cryptate emission (TRACE) technology and the CIS automated assays (CIS UK Ltd., High Wycombe, Bucks, UK). The between day precision of these assays (CV%) was 4.7% for AFP at 11 kU/l and 3.2% for total hCG at 105 800 U/l.

Statistical analysis

Weighted regression analysis was carried out to derive the relationship between marker levels and gestational age. To correct for marker variation with gestational age each value was converted to MoMs from the respective median marker levels in unaffected pregnancies of the same gestational age. Statistical analysis of data was performed using Excel and Astute, a statistical software add-in for Microsoft Excel 5 (DDU

Table 1—Median and range for maternal age, gestational age, fetal crown–rump length, maternal weight and sample storage time in the trisomy 21 group and the control group

	Trisomy 21	Controls
No.	130	959
Maternal age (years)	37 (17–45)	28.8 (15–45)
Gestational age (days)	86 (43–97)	83 (42–97)
Crown–rump length (mm)	61 (42–84)	56 (36–84)
Maternal weight (kg)	64 (44–133)	65 (36–120)
Sample storage time (days)	937 (150–2317)	435 (350–502)

Software, University of Leeds, U.K.) or Analyse-It (Smart Software, Leeds, U.K.). The performance of various marker combinations as potential screening procedures was examined using standard statistical modelling techniques (Royston and Thompson, 1992). Using the observed population parameters for AFP and total hCG and those for other markers detailed in a previous study (Spencer *et al.*, 1999a), a series of 15 000 random MoM values were selected for each marker from within the gaussian distributions of the log MoM of affected and unaffected pregnancies. These values were then used to calculate likelihood ratios for the combinations. The likelihood ratios were then used together with the age-related risk of trisomy 21 in the first trimester (Snijders *et al.*, 1995) to calculate the expected detection rate of affected pregnancies at a fixed false positive rate, in a population with the maternal age distribution of pregnancies in England and Wales (OPCS, 1994).

RESULTS

Table 2 shows the observed and regressed median values for total hCG and AFP. Total hCG MoM and AFP MoM in both the trisomy 21 group and in the control group fitted a gaussian distribution after removal of outliers outside of three standard deviations (number of outliers were seven (0.7%) for total hCG in controls and five (4%) in the trisomy group; for AFP, one (0.1%) in controls and four (3%) in the trisomy group). After \log_{10} transformation, Kolmogorov–Smirnov and Anderson Darling tests showed linearity at the 0.01 probability level for both affected and control populations. Figures 1 and 2 show the probability distributions for both analytes in each study population. Table 3 summarizes the statistical parameters associated with the distributions of the biochemical markers. Correlation of AFP and total hCG in controls and affected populations showed r values of 0.008 and 0.073 respectively. Correlation with free β -hCG and PAPP-A measured previously (Spencer *et al.*, 1999a) and in the additional 40 cases showed a correlation (r) of 0.0847 for free β -hCG and AFP, 0.1859 for PAPP-A and AFP, and 0.4272 for PAPP-A and total hCG in the affected populations. In the unaffected population the correlation between PAPP-A and total hCG was 0.2145.

Figures 3 and 4 show the individual total hCG and AFP MoM's for the trisomy 21 cases. The median MoM for total hCG was 1.31 and this was significantly higher than the control median (MannWhitney U test, $p < 0.0001$). Of these cases, 26/130 (20.0%) were above the 95th centile of normal (1.859 MoM). The median MoM for AFP was 0.82 and this was significantly lower than the control median (MannWhitney U test, $p < 0.0001$). Of these cases, 20/130 (15.4%) were below the 5th centile of normal (0.516 MoM). In the 90 cases which were analysed in our previous study of free β -hCG and PAPP-A (Spencer *et al.*, 1999a) the median MoM in this same group was 2.03 and 0.51 respectively, demonstrating the greater

Table 2—Observed and regressed median levels of AFP and total hCG in unaffected pregnancies

Week	Mean gestation (days)	Median observed AFP (kU/l)	Regressed median AFP (kU/l)	Median observed total hCG (U/l)	Regressed median total hCG (U/l)
6	44	3.1	2.9	71036	72200
7	52	5.2	4.0	87820	88100
8	59	5.7	5.1	104800	100210
9	67	6.2	6.9	114800	125147
10	74	8.4	8.9	84670	82461
11	80	11.1	11.0	79230	72318
12	87	14.1	14.3	63370	62051
13	94	17.5	18.4	53760	53241

shift in median MoM for these two markers compared with total hCG and AFP. Adding to this analysis the 40 additional cases from the Glasgow centre gave an overall median free β -hCG MoM of 2.04 and 0.50 for PAPP-A. The log MoM standard deviation for free β -hCG and PAPP-A was 0.281 and 0.292, these were not significantly different from our original series (Spencer *et al.*, 1999a).

When the observed statistical parameters were used in the mathematical model of a population with a maternal age distribution of pregnancies in England

and Wales, the estimated detection rates using various marker combinations with maternal age at a fixed false positive rate of 5% varied from 31% for AFP and maternal age or total hCG and maternal age, to 40% when both markers were combined with maternal age. If PAPP-A was used (using population data from Spencer *et al.* (1999a), then in combination with total hCG and maternal age, detection rates would increase to 52% compared with the 67% obtained using free β -hCG, PAPP-A and maternal age (Spencer *et al.*, 1999a). We estimated that adding AFP to either of the two marker combinations of total hCG and PAPP-A or free β -hCG, would increase the detection rate by less than 0.5%.

Examination of Figure 4 indicates that the degree of elevation for total hCG MoM in cases of trisomy 21 may be related to the gestational age. When the study group was analysed by gestational age bands a significant difference in median MoM was noted for total hCG but not for AFP. With total hCG the median MoM in trisomy 21 pregnancies less than 70 days was 0.48, rising to 1.13 at 70–83 days and to 1.52 at 84–97 days. The difference between the 70–83 day median and the 84–97 day median was highly significant ($p=0.004$, Mann Whitney U test). The correlation coefficient between \log_{10} total hCG MoM and gestational age in days was 0.444.

When we concentrated our modelling analysis on those cases between 10 and 13 weeks and split them into two groups based on those between 70–83 days and those 84–97 days, the population parameters for total hCG shown in Table 4 were found. Using these population parameters and those for PAPP-A across the 70–97 day gestational period from our previous study (Spencer *et al.*, 1999a), modelled detection rates were 47% at 70–83 days and 60% at 84–97 days, each at a 5% false positive rate.

DISCUSSION

In large-scale studies of first trimester maternal serum (Spencer *et al.*, 1999a), the markers free β -hCG and PAPP-A alone have been shown to identify 33% and 38% of cases of Down syndrome at a 5% false positive rate. When each marker is combined with maternal

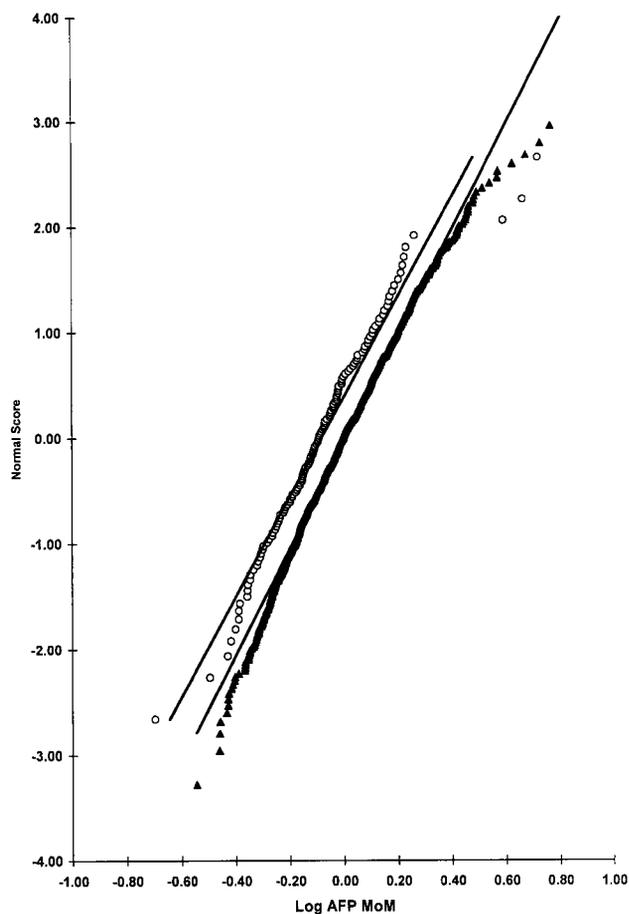


Figure 1—Probability plot of \log_{10} AFP MoM in normal pregnancies (\circ) and Down syndrome pregnancies (\blacktriangle)

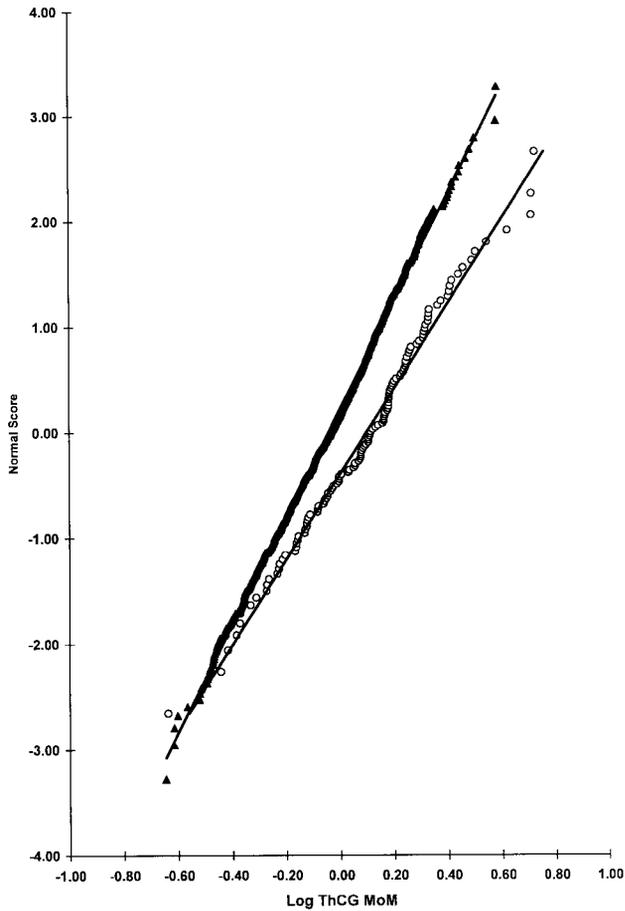


Figure 2—Probability plot of \log_{10} total hCG MoM in normal pregnancies (▲) and Down syndrome pregnancies (○)

age this increases to 46% and 48% respectively, and when combined together in a three parameter algorithm, detection rates for a 5% false positive rate are increased to 67%. Our study of total hCG and AFP shows these marker performances to be inferior to free β -hCG and PAPP-A, with total hCG detecting only 20% of cases and AFP 15% of cases at a 5% false positive rate. When combined with maternal age these rise to 31% for either marker and as a three parameter algorithm with maternal age a detection rate of 40%

Table 3—Statistical parameters for the various marker distributions (as MoM) in trisomy 21 and control pregnancies

	AFP	Total hCG
\log_{10} mean controls	0.008	-0.0044
\log_{10} SD controls	0.1966	0.1959
\log_{10} mean affected	-0.082	0.097
\log_{10} SD affected	0.2111	0.2479
10th centile controls	0.587	0.489
50th centile controls	1.00	1.00
90th centile controls	1.795	1.554
10th centile affected	0.463	0.593
50th centile affected	0.820	1.310
90th centile affected	1.545	2.522

can be achieved. These detection rates appear at odds with those published by Haddow *et al.* (1998), who claimed detection rates of 63% and 60% when maternal age, PAPP-A, and total hCG or free β -hCG were used at a 5% false positive rate. However, as pointed out by Hallahan *et al.* (1998), the observation that total hCG performed better than free β -hCG (29% versus 25% detection rate at a 5% false positive rate) was inconsistent with the distribution parameters provided in the paper, and that on re-analysis the data would show that free β -hCG should have detected 29% compared with 25% for total hCG.

Our current data clearly show that total hCG and AFP are poor markers of trisomy 21 in the first trimester. The median AFP MoM of 0.82 in the Down group is consistent with the summary value of 0.78 in the 16 series (335 cases) summarized in Wald *et al.* (1997). The median total hCG MoM of 1.31 is close to the summary value of 1.29 in the 12 series (352 cases) summarized by Wald *et al.* (1997). However, our data clearly show a gestational age-related increase in median total hCG MoM in cases of trisomy 21. If all of the published series for total hCG are analysed in two groups according to whether the median study gestational age was 11 weeks or older or less than 11 weeks (Table 5), then it is also clear that in general those studies that have claimed total hCG to be useful have higher median MoMs because the study consists predominately of late first trimester samples. The median MoMs in these two similar sized groups (1.199 versus 1.412) show a similar level of difference to our analysis of mid and late first trimester samples.

Temporal changes in median analyte levels across the first and second trimester of affected pregnancies are only now being recognized as large series of data are compiled. It has been known for some time that PAPP-A levels change significantly between the first and second trimester (Berry *et al.*, 1997) but clear demonstration of a gradual change across both periods has only recently been demonstrated (Spencer *et al.*, 1999a). Similarly, changes have recently been demonstrated for trisomy 18 when the reduced PAPP-A levels get progressively lower with advancing gestation (Tul *et al.*, 1999; Spencer *et al.*, 1999b). Free β -hCG demonstrates different temporal changes with the median MoM being highest at 14–16 weeks and gradually falling during the 16–20 (Spencer *et al.*, 1993) and the 10–14 week periods (Spencer *et al.*,

Table 4—Statistical parameters for total hCG distribution in trisomy 21 and control pregnancies split by gestational age

	70 to 83 days	84 to 97 days
\log_{10} mean controls	-0.09909	-0.00509
\log_{10} SD controls	0.1967	0.1995
\log_{10} mean affected	0.019137	0.16983
\log_{10} SD affected	0.1600	0.2253
Number of affected cases	31	89

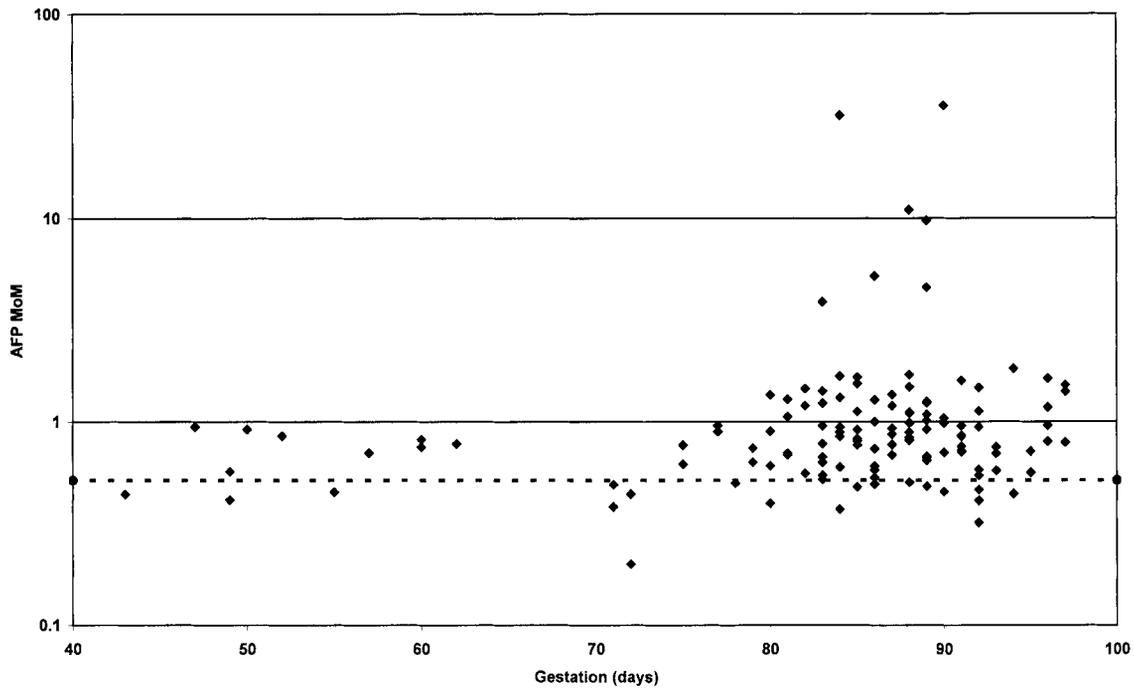


Figure 3—Distribution of AFP MoM with gestational age in Down syndrome cases. Dotted line is the 5th centile of normality

1999a), and falling precipitously prior to 10 weeks (Spencer *et al.* (this article)).

Studies that have suggested that total hCG and PAPP-A can be a useful first trimester screening combination have shown detection rates of 63% (Haddow *et al.*, 1998), very similar to our data for samples collected during the 85–97 day period. Such studies have been biased by the inclusion of samples at the very end of the first trimester. In screening practice

samples will be collected over a much wider gestational range and be matched to the time period that nuchal translucency is valid (73–97 days). We have shown therefore that temporal changes in analyte median MoM in affected cases will have considerable importance and impact on detection rates and will require us to focus screening on a defined optimum period using specifically optimized algorithms (Reynolds *et al.*, 1998).

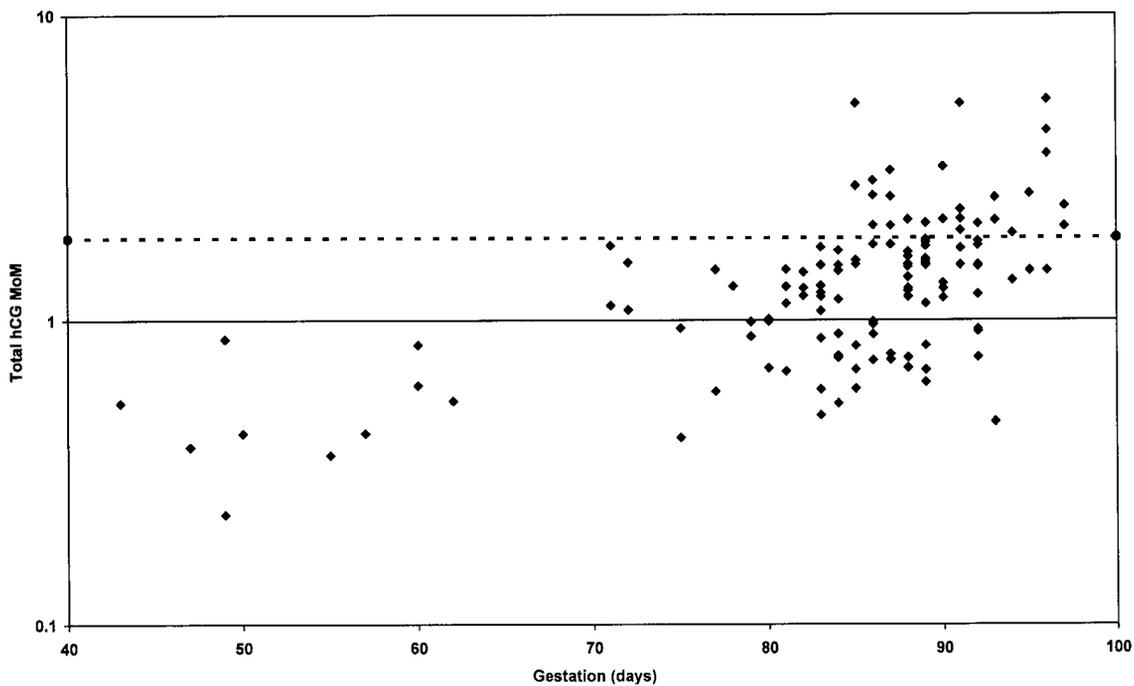


Figure 4—Distribution of total hCG MoM with gestational age in Down syndrome cases. Dotted line is the 95th centile of normality

Table 5—Published studies of total hCG in the first trimester grouped by median gestational age

Study	No. of cases	Median MoM	Weeks' gestation (median and range)
<i>Studies with median gestation <11 weeks</i>			
Brock <i>et al.</i> (1990)	21	1.43	10 (7–14)
Johnson <i>et al.</i> (1991)	11	0.91	10 (9–12)
Kratzer <i>et al.</i> (1991)	17	1.23	10 (9–12)
Hogdall <i>et al.</i> (1992)	14	1.10	10 (9–12)
Van Lith <i>et al.</i> (1992)	24	1.19	10 (9–12)
Aitken <i>et al.</i> (1993)	16	0.97	10 (6–14)
Macintosh <i>et al.</i> (1994)	20	1.40	10 (8–14)
Biagiotti <i>et al.</i> (1995)	41	1.12	10 (8–12)
Wald <i>et al.</i> (1996)	77	1.23	10 (8–14)
Total	241	1.119	
<i>Studies with median gestation ≥11 weeks</i>			
Cuckle <i>et al.</i> (1988)	22	1.10	11 (7–12)
Crandall <i>et al.</i> (1993)	11	1.73	12 (11–14)
Brizot <i>et al.</i> (1995)	41	1.50	11 (10–13)
Haddow <i>et al.</i> (1998)	48	1.54	11 (9–13)
Casals <i>et al.</i> (1999)	32	1.63	12 (10–14)
Spencer <i>et al.</i> (this article)	130	1.31	12 (10–14)
Total	284	1.412	
<i>Other studies with median gestation indeterminate</i>			
Bogart <i>et al.</i> (1989)	6	1.17	(9–11)
Isles <i>et al.</i> (1993)	25	1.35	(8–14)
Kellner <i>et al.</i> (1994)	5	0.90	(8–14)
Forest <i>et al.</i> (1995)	12	1.83	(9–13)
Hallahan <i>et al.</i> (1998)	63	1.3	(10–13)
All studies	636	1.319	

ACKNOWLEDGEMENTS

We acknowledge the support of CIS in providing the reagents and instrumentation to carry out this study.

REFERENCES

- Aitken DA, McGaw G, Crossley JA, Berry E, Connor JM, Spencer K, Macri JN. 1993. First trimester biochemical screening for fetal chromosome abnormalities and neural tube defects. *Prenat Diagn* **13**: 681–689.
- Berry E, Aitken DA, Crossley JA, Macri JN, Connor JN. 1997. Screening for Down syndrome: changes in marker levels and detection rates between first and second trimesters. *Br J Obstet Gynaecol* **104**: 811–817.
- Biagiotti R, Cariati E, Brizzi L, D'Agata A. 1995. Maternal serum screening for Down's syndrome in the first trimester. *Br J Obstet Gynaecol* **102**: 660–662.
- Bogart M, Golbus M, Sorg N, Jones OW. 1989. Human chorionic gonadotropin levels in pregnancies with aneuploid fetuses. *Prenat Diagn* **9**: 379–384.
- Brambati B, Lanzani A, Tului L. 1991. Ultrasound and biochemical assessment of first trimester pregnancy. In *The Embryo: Normal and Abnormal Development and Growth*, Chapman M, Grudzinskas JG, Chard T (eds). Springer-Verlag: New York; 181–194.
- Brizot ML, Snijders RJM, Butler J, Bersinger NA, Nicolaidis KH. 1995. Maternal serum hCG and fetal nuchal translucency thickness for the prediction of fetal trisomies in the first trimester of pregnancy. *Br J Obstet Gynaecol* **102**: 127–132.
- Brock DJH, Barron L, Holloway S, Liston WA, Hillier SG, Seppala M. 1990. First trimester maternal serum biochemical indicators in Down's syndrome. *Prenat Diagn* **10**: 245–251.
- Casals E, Aibar C, Martinez JM, Borrel A, Soler A, Ojuel J, Ballesta AM, Fortuny A. 1999. First trimester biochemical markers for Down syndrome. *Prenat Diagn* **19**: 8–11.
- Crandall BF, Hanson FW, Keener S, Matsumoto M, Miller W. 1993. Maternal serum screening for AFP, uE₃ and hCG between 11 and 15 weeks of pregnancy to detect fetal chromosome abnormalities. *Am J Obstet Gynecol* **168**: 1864–1869.
- Crossley JA, Aitken DA, Berry E, Connor JM. 1994. Impact of a regional screening programme using serum alpha fetoprotein (AFP) and human chorionic gonadotrophin (hCG) on the birth incidence of Down's syndrome in the west of Scotland. *J Med Screen* **1**: 80–83.
- Cuckle HS, Wald NJ, Barkai G, Fuhrmann W, Altland K, Brambati B, Knight G, Palomaki G, Haddow JE, Canick J. 1988. First trimester biochemical screening for Down's syndrome. *Lancet* **ii**: 851–852.
- Cuckle HS, Ellis AR, Seth J. 1995. Provision of screening for Down's syndrome. *Br Med J* **311**: 512.
- Forest JC, Masse J, Rousseau F, Moutquin JM, Brideau NA, Belanger M. 1995. Screening for Down syndrome during the first and second trimesters: impact of risk estimation parameters. *Clin Biochem* **28**: 443–449.
- Goodburn SF, Yates JRW, Raggatt PR. 1994. Second trimester maternal serum screening using alpha-fetoprotein, human chorionic gonadotrophin and unconjugated oestriol: experience of a regional programme. *Prenat Diagn* **14**: 391–402.
- Haddow JE, Palomaki GE, Knight GJ, Williams J, Miller WA, Johnson A. 1998. Screening of maternal serum for fetal Down's syndrome in the first trimester. *N Engl J Med* **338**: 955–961.
- Hallahan T, Krantz D, Khabbaza E, Orlandi F, Rossi C, Curcio P, Macri S, Larsen J, Buchanan P, Macri J. 1998. First trimester maternal serum screening for Down syndrome: intact hCG versus free beta hCG. *Am J Hum Genet* **63** (Suppl): A164.
- Hogdall CK, Hogdall EVS, Arends J, Noorgard-Pedersen B, Smidt-Jensen S, Larsen SO. 1992. CA-125 as a maternal serum marker for Down's syndrome in the first and second trimesters. *Prenat Diagn* **12**: 223–227.
- Isles RK, Sharma K, Wathen NC, Campbell J, Ward H, Muller F, Grudzinskas JG, Chard T. 1993. hCG, free subunit and PAPP-A composition of maternal serum in normal and Down's syndrome

- pregnancies. Fourth Conference: *Endocrinology and Metabolism in Human Reproduction*. Royal College of Obstetrics and Gynaecology: London.
- Johnson A, Cowchock FS, Darby M, Wapner R, Jackson LG. 1991. First trimester maternal serum AFP and hCG in aneuploid pregnancies. *Prenat Diagn* **11**: 443–450.
- Kellner LH, Weiss RR, Weiner Z, Neur M, Martin G. 1994. Early first trimester maternal serum AFP, uE₃ hCG and free beta-hCG measurements in unaffected and affected pregnancies with fetal Down syndrome. *Am J Hum Genet* **55** (Suppl): A281.
- Kratzer PG, Golbus MS, Monroe SE, Finkelstein DE, Taylor RN. 1991. First trimester aneuploidy screening using serum human chorionic gonadotropin (hCG), free alpha-hCG and progesterone. *Prenat Diagn* **11**: 751–765.
- Macintosh MCM, Iles R, Teisner B, Sharma K, Chard T, Grudzinskas JG, Ward RH, Muller F. 1994. Maternal serum hCG and PAPP-A markers for fetal Down syndrome at 8–14 weeks. *Prenat Diagn* **14**: 203–208.
- Macri JN, Spencer K. 1996. Towards the optimal protocol for Down's syndrome screening. *Am J Obstet Gynecol* **174**: 1668–1669.
- Office of Population Censuses and Surveys. 1994. *Birth Statistics 1986–1994*. Series FM1, Nos 13–21. HMSO: London.
- Palomaki GE, Knight GJ, McCarthy JE, Haddow JE, Donhove JM. 1997. Maternal serum screening for Down syndrome in the United States: a 1995 survey. *Am J Obstet Gynecol* **176**: 1046–1051.
- Reynolds TM, Dunstan F, Nix B, Williams K, Crossley J, Holding S, Krantz D, Wright D, Bray I, Spencer K. 1998. Combining ultrasound and biochemistry in first trimester screening for Down's syndrome. Response to Wald and Hackshaw. *Prenat Diagn* **18**: 511–515.
- Royston P, Thompson SG. 1992. Model based screening for risk with application to Down's syndrome. *Stats Med* **11**: 257–268.
- Snijders RJM, Sebire NJ, Nicolaides KH. 1995. Maternal age and gestational age specific risk for chromosomal defects. *Fetal Diagn Ther* **10**: 356–357.
- Snijders RJM, Noble P, Sebire N, Souka A, Nicolaides KH, for the Fetal Medicine Foundation First Trimester Screening Group. 1998. UK multicentre project on assessment of risk of trisomy 21 by maternal age and fetal nuchal translucency thickness at 10–14 weeks of gestation. *Lancet* **351**: 343–346.
- Spencer K. 1997. hCG and its subunits in first trimester Down syndrome screening. In *Screening for Down Syndrome in the First Trimester*, Grudzinskas JG, Ward RHT (eds). RCOG Press: London; 117–131.
- Spencer K. 1999a. Second trimester prenatal screening for Down's syndrome using alpha-fetoprotein and free beta hCG: a seven-year review. *Br J Obstet Gynaecol* **106**: 1287–1293.
- Spencer K. 1999b. One stop clinic for assessment of risk for fetal abnormalities. *Downs Screening News* **6**: 10.
- Spencer K, Macri JN, Aitken DA, Connor JM. 1992. Free beta-hCG as a first trimester marker for fetal trisomy. *Lancet* **339**: 1480.
- Spencer K, Macri JN, Anderson RW, Aitken DA, Berry E, Crossley JA, Wood PJ, Coombes EJ, Stroud M, Worthington DJ, Doran J, Barbour H, Wilmot R. 1993. Dual analyte immunoassay in neural tube defect and Down's syndrome screening: results of a multicentre clinical trial. *Ann Clin Biochem* **30**: 394–401.
- Spencer K, Aitken DA, Crossley JA, McGaw G, Berry E, Anderson R, Connor JM, Macri JN. 1994. First trimester biochemistry screening for trisomy 21: the role of free beta hCG, alpha fetoprotein and pregnancy associated plasma protein A. *Ann Clin Biochem* **31**: 447–454.
- Spencer K, Souter V, Tul N, Snijders R, Nicolaides KH. 1999a. A screening program for trisomy 21 at 10–14 weeks using fetal nuchal translucency, maternal serum free β -human chorionic gonadotropin and pregnancy associated plasma protein-A. *Ultrasound Obstet Gynecol* **13**: 231–237.
- Spencer K, Crossley JA, Green K, Worthington DJ, Brownbill K, Aitken DA. 1999b. Second trimester levels of pregnancy associated plasma protein-A in cases of trisomy 18. *Prenat Diagn* **19**: 1127–1134.
- Tul N, Spencer K, Noble P, Chan C, Nicolaides K. 1999. Screening for trisomy 18 by fetal nuchal translucency and maternal serum free β -hCG and PAPP-A at 10–14 weeks of gestation. *Prenat Diagn* **19**: 1035–1042.
- Van Lith JMM. 1992. First trimester maternal serum hCG as a marker for fetal chromosomal disorders. *Prenat Diagn* **12**: 495–504.
- Wald NJ, Kennard A, Densem JW, Cuckle HS, Chard T, Butler L. 1992. Antenatal maternal serum screening for Down's syndrome: results of a demonstration project. *BMJ* **305**: 391–394.
- Wald NJ, George L, Smith D, Densem JW, Petterson K, on behalf of the International Prenatal Screening Research Group. 1996. Serum screening for Down's syndrome between 8 and 14 weeks of pregnancy. *Br J Obstet Gynaecol* **104**: 407–412.
- Wald NJ, Kennard A, Hackshaw A, McGuire A. 1997. Antenatal screening for Down's syndrome. *J Medical Screening* **4**: 181–246.